

Fibrinogen in traumatic haemorrhage: A narrative review

Winearls, James; Campbell, Don; Hurn, Catherine; Furyk, Jeremy; Ryan, Glenn; Trout, Melita; Walsham, James; Holley, Anthony; Shuttleworth, Megan; Dyer, Wayne; Keijzers, Gerben; Presneill, Jeff; Fraser, John F.; Wullschleger, Martin

Published in:
Injury

DOI:
[10.1016/j.injury.2016.12.012](https://doi.org/10.1016/j.injury.2016.12.012)

Licence:
CC BY-NC-ND

[Link to output in Bond University research repository.](#)

Recommended citation(APA):

Winearls, J., Campbell, D., Hurn, C., Furyk, J., Ryan, G., Trout, M., Walsham, J., Holley, A., Shuttleworth, M., Dyer, W., Keijzers, G., Presneill, J., Fraser, J. F., & Wullschleger, M. (2017). Fibrinogen in traumatic haemorrhage: A narrative review. *Injury*, 48(2), 230-242. <https://doi.org/10.1016/j.injury.2016.12.012>

General rights

Copyright and moral rights for the publications made accessible in the public portal are retained by the authors and/or other copyright owners and it is a condition of accessing publications that users recognise and abide by the legal requirements associated with these rights.

For more information, or if you believe that this document breaches copyright, please contact the Bond University research repository coordinator.

Fibrinogen in Traumatic Haemorrhage: A Narrative Review

Dr James Winearls, BSc (Hons), MBBS, MRCP, FCICM, Consultant Intensivist, Gold Coast University Hospital. Senior Lecturer, School of Medicine, University of Queensland, Senior Lecturer, School of Medical Sciences, Griffith University.

Corresponding Author. james.winearls@health.qld.gov.au and
james.winearls@gmail.com

Dr Don Campbell, MBBS, FACEM, Consultant Emergency Physician and Deputy Director Trauma, Gold Coast University Hospital. don.campbell@health.qld.gov.au

Dr Catherine Hurn, MBBS, FACEM, Consultant Emergency Physician, Royal Brisbane and Women's Hospital. Senior Lecturer, School of Medicine, University of Queensland. catherine.hurn@health.qld.gov.au

A/Prof Jeremy Furyk, MBBS, FACEM, Consultant Emergency Physician, Director of Emergency Research, Townsville Hospital. jeremy.furyk@health.qld.gov.au

Dr Glenn Ryan, MBBS, FACEM, Consultant Emergency Physician, Princess Alexandra Hospital. glenn.ryan@health.qld.gov.au

Dr Melita Trout, MBBS, FCICM, Consultant Intensivist, Townsville Hospital.
melita.trout@health.qld.gov.au

Dr James Walsham, MBChB, MRCP FCICM, Consultant Intensivist and Director of Intensive Care Research, Princess Alexandra Hospital. Senior Lecturer, School of Medicine, University of Queensland. james.walsham@health.qld.gov.au

Dr Anthony Holley, MBBCh, FACEM, FCICM, Consultant Intensivist, Royal Brisbane and Women's Hospital. Senior Lecturer, School of Medicine, University of Queensland. anthony.holley@health.qld.gov.au

Ms Megan Suttleworth, BHSc, MMedRes, Menzies Health Institute Queensland, Griffith University. m.shuttleworth@griffith.edu.au

Dr Wayne Dyer, PhD, Senior Research Fellow, Australian Red Cross Blood Service. W.Dyer@redcrossblood.org.au

A/Prof Gerben Keijzers, MSc, MBBS, PhD, FACEM. Consultant Emergency Physician, Gold Coast University Hospital. Senior Lecturer, School of Medicine, Bond University. Senior Lecturer, School of Medical Sciences, Griffith University. gerben.keijzers@health.qld.gov.au

Professor Jeff Presneill, MBBS, PhD, FCICM, Consultant Intensivist, Director of Intensive Care Research, Royal Melbourne Hospital. jeffrey.presneill@health.qld.gov.au

Professor John Fraser, MBChB, PhD, FRCP (Glas), FRCA, FFARCSI, FCICM,
Consultant Intensivist, Director of Critical Care Research Group, The Prince Charles
Hospital and University of Queensland. john.fraser@health.qld.gov.au

A/Prof Martin Wulschleger, MBBS, MD, PhD, FRACS, Director of Trauma, Gold
Coast University Hospital. Senior Lecturer, School of Medical Sciences, Griffith
University. martin.wulschleger@health.qld.gov.au

Abstract:

Haemorrhage in the setting of severe trauma is associated with significant morbidity and mortality. There is increasing awareness of the important role fibrinogen plays in traumatic haemorrhage. Fibrinogen levels fall precipitously in severe trauma and the resultant hypofibrinogenaemia is associated with poor outcomes. Hence, it has been postulated that early fibrinogen replacement in severe traumatic haemorrhage may improve outcomes, although, to date there is a paucity of high quality evidence to support this hypothesis. In addition there is controversy regarding the optimal method for fibrinogen supplementation. We review the current evidence regarding the role of fibrinogen in trauma, the rationale behind fibrinogen supplementation and discuss current research.

Abbreviations: A5 (A10) = amplitude of clot firmness 5 (10) minutes after CT, BW = body weight, CFT = clot formation time, Cryo = cryoprecipitate, CT = clotting time, DCR = damage control resuscitation, ED = Emergency Department, FC = fibrinogen concentrate, FibC = Clauss Fibrinogen, FFP = fresh frozen plasma, GCUH = Gold Coast University Hospital, Queensland, Australia, Hb = Hemoglobin, iCa = ionized Calcium, ICU = intensive care unit, MCF = maximum clot firmness, MHP = major haemorrhage protocol, OR = operating room, rPT = rapid Prothrombin Time, POC = Point of Care, PRBC = packed red blood cells, PT = Prothrombin Time SLT = standard laboratory test, Temp = core temperature, TIC = trauma-induced coagulopathy, TXA = tranexamic acid, VHA = viscoelastic hemostatic assays

Key Words: Trauma, Coagulopathy, Fibrinogen, Cryoprecipitate, Fibrinogen Concentrate, Massive Transfusion, Viscoelastic Haemostatic Assays.

Introduction:

Trauma is a leading cause of death worldwide in individuals aged 18-39 years and represents a major global health concern (1). Despite advances in trauma management, a significant proportion of these deaths are secondary to haemorrhage and preventable (2) (3) (4). In patients where surgical haemorrhage control is achieved, subsequent morbidity and mortality is often attributed to coagulopathy complicated by organ failure due to the effects of major haemorrhage and large volume blood product transfusion (5) (6) (7). Death related to major haemorrhage is potentially preventable and represents a target for mortality reduction strategies. There is increasing awareness regarding the critical role of fibrinogen in traumatic

haemorrhage. The objectives of this review are to examine the available evidence regarding fibrinogen in severe trauma and explore its potential role in management.

Methods:

A literature search was performed on major databases including PubMed, Medline, Embase, Web of Science and ClinicalTrials.gov. Search terms included “fibrinogen”, “trauma”, “transfusion protocols”, “massive transfusion”, “cryoprecipitate”, “fibrinogen concentrate”, “viscoelastic testing”, “ROTEM” and “TEG”. Titles and abstracts were reviewed and full text articles retrieved for inclusion if considered relevant. Additional articles were identified from reference lists of identified articles. Database searches were supplemented by private files/collections of the authors, and a grey literature search using Google scholar. Results are presented in a narrative form.

Role of Fibrinogen in Trauma Induced Coagulopathy:

Severe trauma may be complicated by a unique, complex and multifactorial coagulopathy – Trauma Induced Coagulopathy (TIC); of which the exact pathophysiological mechanisms are yet to be elucidated (8) (9) (10) (11). TIC is characterized by reduced clot strength related to hypo/dysfibrinogenaemia, platelet dysfunction, hyperfibrinolysis and endothelial dysfunction (12). Central to the proposed mechanism is the effect of direct tissue injury and hypoperfusion on the endothelium resulting in systemic anticoagulation and hyperfibrinolysis (13). TIC may subsequently be exacerbated by acidosis, hypothermia, haemodilution and factor consumption (14) While the exact mechanisms are debated, patients with TIC do

certainly have significantly increased transfusion requirements and mortality (15) (16).

The role of fibrinogen in maintaining effective haemostasis is widely accepted (17). Fibrinogen is a glycoprotein synthesised in the liver and is the key final component of the clotting cascade; forming fibrin – an insoluble protein that is the foundation of a stable clot (18). Fibrinogen is cleaved by Thrombin to fibrin monomers, which are polymerised and subsequently stabilised by activated Factor XIII to form a fibrin clot (19). Fibrinogen is also fundamental in the aggregation of activated platelets through the glycoprotein IIb/IIIa receptors. The healthy adult has a plasma fibrinogen concentration of between 2 to 4g/L (20).

In traumatic haemorrhage, there is increasing evidence supporting the important role fibrinogen plays in effective clot formation. Lower levels of fibrinogen and increased fibrinogen breakdown are key features of TIC (21) (22). Fibrinogen is the first factor to fall below reference values during bleeding and in trauma reaches critically low levels earlier than any other coagulation factors (23). This has been demonstrated in both the pre-hospital environment and early after arrival in the trauma unit prior to large volume fluid resuscitation (24) (25). In a number of studies the degree of hypofibrinogenaemia is strongly associated with injury severity (22) (26). This suggests fibrinogen deficiency is innately related to the primary and secondary physiological insults induced by severe trauma. Subsequently, further hypofibrinogenaemia occurs as a consequence of major blood loss, consumption, dilution, acidosis and hypothermia (27) (28) (29) (30). In addition, the fibrin strands that form in a low fibrinogen environment are more susceptible to fibrinolysis and a

number of studies report hypofibrinogenaemia in the presence of hyperfibrinolysis (31) (32) (33). Hyperfibrinolysis in the setting of severe trauma is a central component of TIC and although relatively rare is associated with poor outcomes (34) (35) (36). The association between hypofibrinogenaemia and worse outcomes in severe trauma has been well demonstrated although to date the pathophysiological mechanisms remain incompletely understood (22) (37) (38).

Major Haemorrhage Protocols (MHP):

Major Haemorrhage Protocols utilised by many trauma centres are activated once significant haemorrhage is suspected. In addition to rapid surgical control of haemorrhage, the MHP involves the empiric and early delivery of a predefined fixed-ratio transfusion of blood products (Plasma, Platelets and PRBC), in an attempt to ameliorate the coagulopathy associated with major haemorrhage and large volume blood transfusion (39) (40). MHP have been implemented in response to studies showing that in massive transfusion the inadequate replacement of coagulation factors is associated with poor outcomes (41). Whilst it is clear that the institution of a MHP does improve outcomes, the optimal ratio of blood components to PRBC remains the focus of considerable debate (42) (43) (44) (45) (46).

The PROMMTT study provided initial data to support a high product to PRBC transfusion ratios in trauma MHP (1:1:1 Plasma:Platelets:PRBC) (47). The survival advantage postulated to be due to earlier replacement of consumed factors mitigating the effect of TIC and that the replacement physiologically more closely approximates to what is being lost. However, even in high ratios the delivered

replacement is potentially dilute in terms of factors (39). The subsequent Pragmatic Randomized Optimal Platelet and Plasma Ratios (PROPPR) trial reported no difference in survival between a 1:1:1 or 1:1:2 transfusion strategy but more patients in the 1:1:1 group achieved haemostasis and fewer died from exsanguination (48). A number of studies have reported that in patients receiving higher ratios there is an increase in the amount of plasma transfused and an increased incidence of transfusion-related adverse events without a survival benefit, however, these concerns were not confirmed in the PROPPR trial (49) (50) (51). Time to delivery of product replacement in PROPPR was rapidly achieved without significant wastage but this is dependent on having thawed product available 24 hours a day in either the trauma unit or blood bank (52). The impressive delivery of blood component therapy as described in the PROPPR trial may be difficult to translate into routine clinical practise (53). A recent study from Stanworth et al. reports widespread variations in patterns of blood product delivery across a number of trauma centres, with few patients receiving an 'optimal' product ratio (54).

Fibrinogen Replacement as part of a MHP:

Hypofibrinogenaemia after severe trauma is associated with increased risk of both massive transfusion and mortality (37) (55). It is postulated that early fibrinogen replacement may be efficacious in correction of coagulopathy, assist in haemorrhage control and decrease transfusion requirements (37) (56) (57) (58). Two studies from the military indicate that maintaining higher fibrinogen levels as part of a MHP is associated with improved survival (59) (60).

Empiric and early fixed ratio delivery of specific fibrinogen containing products is not standard practice in the majority of MHP. Whilst plasma does contain fibrinogen it is in a concentration and volume that is potentially too dilute in terms of adequacy of replacement. The mean fibrinogen concentration in plasma is approximately 2g/L and large volumes of plasma (+/- 30ml/Kg) are required to adequately supplement fibrinogen (61). In most MHP the transfusion of additional fibrinogen in the form of cryoprecipitate usually occurs only late in the protocol and in response to low plasma fibrinogen levels as measured by laboratory tests of plasma fibrinogen. There are inherent problems with this approach resulting in significant delays to effective administration of fibrinogen. In addition there is significant debate regarding at what level of plasma fibrinogen should trigger additional fibrinogen replacement. The majority of current guidelines would suggest that plasma fibrinogen levels of < 1 – 1.5g/l should trigger additional fibrinogen, however, this is not based on solid evidence (62) (63). Hagemo et al, have demonstrated an increased mortality in trauma patients with fibrinogen levels below 2.29g/l (22). In severe trauma fibrinogen levels should potentially be maintained at levels higher than currently recommended. The 2016 European Trauma Guidelines recommend fibrinogen supplementation if thromboelastometric signs of functional fibrinogen deficiency or plasma fibrinogen levels < 1.5 – 2g/l (64).

Fibrinogen can be replaced with: fibrinogen concentrate (FC), cryoprecipitate or plasma, each containing different amounts of fibrinogen: 20g/L, 8-16g/l and 2g/L, respectively, and therefore different volumes are required to achieve replacement (65). A study by Rourke et al. (37) demonstrated that standard protocol driven transfusion ratios were ineffective in maintaining fibrinogen levels and the addition of

cryoprecipitate was required. Khan et al. (66) showed that high dose plasma transfusion does not correct TIC and coagulation parameters only improve with plasma, cryoprecipitate and platelet transfusion with a combined high fibrinogen load. Chambers et al. (67) report that 1:1:1 MHP does not affect the frequency or duration of coagulopathy as measured by standard coagulation tests and hypofibrinogenaemia was almost always the first abnormality detected.

The PROMMTT study described wide variability in cryoprecipitate transfusion practices in 10 American Level 1 trauma centres; in those patients receiving cryoprecipitate the median time to transfusion was 2.7 hours and the majority of those patients who died of haemorrhage did not receive cryoprecipitate (68). These findings are supported by a recently published study from the UK investigating transfusion practices in traumatic haemorrhage - the median time to delivery of cryoprecipitate as part of a MHP was more than 2 hours and almost 50% of patients did not receive cryoprecipitate as part of their initial resuscitation (54). This is concerning, considering that the median time to death from haemorrhage is reported to be approximately 2.6 hours (48) (54).

The CRYOSTAT study, recently published by Curry et al. (69), demonstrated that the early administration of cryoprecipitate as part of a MHP is feasible in trauma patients. 85% of patients randomised to the Cryo arm received cryoprecipitate within 90 minutes and the median time to transfusion was 60 minutes. Additionally, fibrinogen levels were consistently higher throughout active haemorrhage in the Cryo arm.

The ideal trauma MHP remains elusive and a number of key questions remain unanswered. Amongst these are: 1) Which laboratory tests and what triggers should be used to guide resuscitation 2) What is the impact of early fibrinogen replacement on haemostasis and clinical outcomes (46).

Assessment of TIC: Standard Laboratory Tests (SLT)

The diagnosis of TIC is conventionally made using prothrombin time (PT) and activated partial thromboplastin time (APTT). These tests, performed on platelet poor plasma were developed to determine single factor deficiencies and effects of anticoagulant therapy. They are poor predictors of bleeding in trauma and due to time delay to result availability fail to provide contemporary information (70) (71). Neither the PT nor APTT give an indication of the fibrinogen contribution to clot strength or quality. There are number of laboratory test utilised to quantify plasma fibrinogen levels, the two most common are; Clauss Fibrinogen (FibC) and Prothrombin Time (PT) Derived Fibrinogen. In the FibC test, concentrated thrombin is added to dilute plasma, converting fibrinogen to fibrin and the clotting time is inversely proportional to the amount of fibrinogen. In the PT-Derived Fibrinogen the difference between baseline and maximum turbidity is proportional to fibrinogen concentration (72) (73). The tests should not be used interchangeably as there can be significant discrepancies between PT-derived fibrinogen and FibC (74). There can be significant variations in reproducibility and consistency in fibrinogen levels between laboratories utilising either test due to differences in; type of analyser, read out method (photo-optical or electromechanical), software and brand of assay used (75). The use of artificial colloids can also falsely elevate the fibrinogen level

reported by both tests (72) (76) (77). However, this is unlikely to be of major concern in Australia, where the use of artificial colloids is not usual standard practice. The major limitation in utilising either test for assessment of fibrinogen concentration in severe trauma is the time delay to result availability, which can be greater than 60 minutes (78). An emergency haemorrhage panel utilising standard laboratory tests (including fibrinogen) with modifications to sample centrifuging, assays and calibration ranges is reported to be available in approximately 20 minutes (79). However, this is not routinely available in Australia and may be difficult to implement into routine clinical practice.

Assessment of TIC: Viscoelastic Haemostatic Assays (VHA):

An alternative to SLT, utilises viscoelastic haemostatic assays (VHA) to rapidly identify coagulation defects and potentially guide targeted interventions (80) (81) (82) (83) (84). Two commercially available devices: TEG® (Haemonetics, Braintree, MA, USA) and ROTEM® (TEM International GmbH, Munich, Germany) are in widespread use; neither is superior and utilisation differs geographically (85) (86) (87). In our institutions the ROTEM® device is utilised; some information is summarised in Figures 1 and 2. VHA measure clot formation up to and including fibrinolysis in contrast to SLT, which document the beginning of fibrin formation when only 5% of total thrombin has been generated. VHA provide information regarding time to clot formation, clot strength and clot lysis; enabling different components of the coagulation cascade and their respective contribution to the clot kinetics to be assessed (85) (88) (89).

VHA have higher sensitivity for the detection of traumatic coagulopathy and provide results more rapidly than SLT (78) (90) (91) (92). In a study by Holcomb et al. (93) involving almost 2000 patients, Thromboelastography (TEG) was found to be superior to SLT across a number of parameters. The use of point of care (POC) rapid PT (rPT) devices can be utilised as an initial tool to assess coagulopathy but their utility remains controversial. A recent paper by Goodman et al, has suggested that POC rPT can be utilised as an alternative to r-TEG, reporting that rPT is cheaper and faster than r-TEG with similar diagnostic accuracy (94). In contrast, Davenport et al, report that although available quickly, the point of care rPT results are inaccurate with significant discrepancies to laboratory PT (78). The rPT may give an indication of the development of TIC and risk of subsequent massive transfusion but neglects the contribution of fibrinogen to clot strength (71). VHA have been incorporated into a number of trauma management guidelines and a number of trauma centres utilise targeted protocols in addition to or in place of fixed ratio MHP (64) (95) (96) (97). Inaba et al, have recently published a consensus statement based on expert opinion and extensive systematic literature review regarding VHA guided treatment triggers for blood product transfusion in severe trauma (81).

However, despite the growing evidence base to support the use of VHA in guiding blood product therapy in traumatic haemorrhage, data from randomised controlled studies is limited (57) (96) (98) (99) (100). A recent Cochrane Review concluded that there is an expanding evidence base that the application of VHA guided transfusion strategies can improve morbidity in bleeding patients (101). However, the quality of studies was low, the majority of trials were in cardiac surgical patients and further high quality studies in acute haemorrhage are required.

A recently published single centre randomised controlled trial reported significant reduction in blood product transfusion rates and improved survival with a TEG guided MHP (97). The multi-centre, prospective randomised controlled iTACTIC Trial (NCT:02593877) investigating the use of VHA in traumatic haemorrhage is currently recruiting in Europe. Patients randomised to the intervention arm will receive MHP resuscitation (1:1:1) with subsequent VHA (ROTEM® or TEG®) guided blood product and pro-coagulant factor administration; the control arm will receive the same 1:1:1 MHP resuscitation with subsequent blood product and pro-coagulant transfusion guided by standard laboratory tests. The primary outcome is proportion of patients alive and free of major haemorrhage at 24 hours. The trial will recruit about 400 patients and is aiming to complete at the end of 2017.

It has been suggested that there can be quality control and standardisation issues associated with point of care viscoelastic testing assays; with variability in test results between different devices, operators and centres (102) (103). It is important that institutions operating viscoelastic devices in the point of care setting should be involved in external quality assurance programmes (104) (105) (106).

Rapid Fibrinogen Assessment Utilising VHA:

Specific assays in both ROTEM® and TEG® are available to rapidly assess fibrinogen contribution to clot strength. In both assays the clot kinetics are assessed in the presence of a platelet inhibitor; FIBTEM (ROTEM®) – Cytochalasin D and Functional Fibrinogen [FF] (TEG®) - Abciximab. The FIBTEM and FF assays can be

utilised to rapidly identify those patients in whom hypo/dysfibrinogenaemia is contributing to on-going haemorrhage (55) (95) (107) (108). The FIBTEM and FF correlate well with standard laboratory measurements of fibrinogen concentration in a number of clinical situations (109) (110) (111) (112) (113).

There is good quality evidence to support the use of the FIBTEM assay as a marker of TIC and in predicting massive transfusion in severely injured trauma patients (90). There is increasing evidence to support the use of the FIBTEM assay as a strategy to decrease blood product transfusion in a variety of clinical settings (98) (114) (115) (116) (117). However, these findings have not been confirmed by high quality studies in the severely bleeding trauma patient.

To optimize capacity to correct coagulopathy rapidly, clot firmness amplitude results obtained at five minutes after clot formation (A5) can be used (55) (107) (118). A5 results have been found to correlate very well with maximum clot firmness (MCF) results in a number of clinical settings (118). Rapidly available real time results potentially permit targeted fibrinogen supplementation to those patients that need it rather than in a fixed-ratio MHP manner or in response to standard laboratory tests. The approach to fibrinogen replacement in our institution is demonstrated in Figures 3 and 4.

Cryoprecipitate for Fibrinogen Replacement:

Cryoprecipitate has been in use for more than 50 years and was originally developed as a treatment for patients with Haemophilia A (119). Each unit of cryoprecipitate is

prepared from 1 unit of FFP; thawed at 1-6°C, centrifuged to remove the excess cryodepleted plasma, re-suspended in 30-40ml of residual plasma and refrozen at -18°C. Although each unit of cryoprecipitate contains a high concentration of fibrinogen due to the small volume, the process recovers only 30% of fibrinogen from the plasma unit. In addition cryoprecipitate contains the other clotting factors - FVIII, vWF and FXIII. Cryoprecipitate is now almost exclusively used to replace fibrinogen in patients with acquired hypofibrinogenaemia – often in the setting of critical bleeding (120).

Cryoprecipitate use in severe trauma accounts for up to 30% of all units transfused (121) (122). There is widespread variability in the recommended dose of cryoprecipitate (ranging from 10 to 20 units) and little conformity between professional institutions (62). There is little high level evidence to support these dosing recommendations and reasons for this variability are twofold. Firstly, the concentration of fibrinogen in cryoprecipitate varies significantly between countries and institutions, ranging from 3-30g/l (62). In a Canadian study, cryoprecipitate units prepared in the same institution had fibrinogen concentrations ranging from 3.2 to 8.2g/l (123). The majority of regulatory authorities state that each unit of cryoprecipitate should contain at least 150mg of fibrinogen. Secondly and directly linked to the variability in fibrinogen per unit of cryoprecipitate, is the variable dose response to cryoprecipitate as determined by plasma fibrinogen levels. It is reported that the transfusion of 10U cryoprecipitate should increase the plasma fibrinogen by approximately 1g/l (124). However, in the trauma setting it has been reported that a dose of approximately 9U cryoprecipitate resulted in a mean increase in plasma fibrinogen of only 0.55g/l (125). The majority of guidelines recommend dosing of

cryoprecipitate in response to low plasma fibrinogen levels. This is impractical in the setting of severe bleeding where delays to effective transfusion of cryoprecipitate are compounded by the time taken to prepare and procure the requested units. A number of papers report the 'inappropriate' transfusion of cryoprecipitate i.e. not given in response to plasma fibrinogen levels (121) (122) (125) (126). It is likely that the delays in obtaining plasma fibrinogen results combined with time to prepare requested units, results in clinicians empirically ordering and transfusing cryoprecipitate on clinical grounds rather than as per published guidelines. The use of cryoprecipitate in traumatic haemorrhage varies widely between institutions and there are often significant delays to effective transfusion (54). The CRYOSTAT study has demonstrated that it is possible to transfuse cryoprecipitate early and empirically as part of a MTP, however, the median time to administration was still 60 minutes (69).

Cryoprecipitate (for fibrinogen replacement) has been withdrawn from use in many European countries due to safety concerns and has been replaced with Fibrinogen Concentrate (127) (128). A standard dose of cryoprecipitate is sourced from multiple donors, therefore potentially increasing the risk of pathogen transmission and transfusion related adverse events (129).

Cryoprecipitate is widely accepted as the standard of care for fibrinogen supplementation in severe haemorrhage; however, there is a lack of good quality evidence to support this strategy. It has been suggested, in view of the fact that cryoprecipitate is not virus-inactivated, is dosed in multiple units, with a lack of quality evidence to support its use and that there are potentially safer alternatives

available, it is unlikely that regulatory approval would be granted for its use today (62) (128).

Fibrinogen Concentrate for Fibrinogen Replacement:

There are a number of theoretical advantages to the use of FC; reduction in volume required, standard dose per vial, lack of variability in fibrinogen concentration, no requirement for ABO compatibility matching, viral inactivation, stored at room temperature, easily reconstituted and administered. However, in severe trauma there are no robust clinical trials demonstrating a survival or cost effectiveness benefit to the use of FC compared to cryoprecipitate (21) (63) (130) (131) (132).

There is increasing evidence supporting the important role of fibrinogen and the use of FC in other clinical situations with severe haemorrhage – cardiac surgery, obstetric haemorrhage and general surgery (133) (134) (135) (136) (137). Recent systematic reviews on the management of major haemorrhage and the use of fibrinogen supplementation suggest potential positive benefits but conclude more research is required (21) (138) (139). A Cochrane review evaluating the effectiveness of FC in severe haemorrhage reported six trials of moderate quality that were underpowered for mortality benefit detection but did demonstrate reduction in allogeneic transfusion requirements (140).

There is expanding observational evidence to support the use of FC in the setting of severe trauma and a number of studies have reported; increased clot strength, reduction in blood loss, reduced transfusion of allogenic blood products and reduction in mortality in patients treated with FC (96) (57) (98) (99) (141). Although

promising, the majority of publications are observational or retrospective cohort studies and do not provide high level evidence to support FC use in severe trauma.

A major concern regarding early fibrinogen replacement utilising FC is the potential for subsequent thromboembolic complications. This is of particular concern in severely traumatised patients who are at significant risk of this complication. A number of animal studies using models of traumatic coagulopathy provided initial safety data in favour of FC (142) (143) (144). Subsequently, a recently published pharmaco-vigilance study suggests that FC is not associated with increased thromboembolic complications; data from over 2.5 million grams of FC (approximately 600,000 standard doses of 4g) distributed over a 27 year period reported possible thromboembolic events in 28 cases (1 per 93,300 grams or 1 per 23,300 doses) (145). A comprehensive systematic review evaluating FC use in the perioperative setting concluded that there was no significant increase in thrombotic events in FC treated patients (146). Schochl et al. report in a cohort of severely injured trauma patients, that even after large doses of FC, subsequent plasma levels of fibrinogen did not exceed normal expected ranges and that there was no increased risk of thromboembolic complications (96).

Fibrinogen dosing:

The optimum dosing schedule of fibrinogen is controversial with widespread variability in recommendations from different professional bodies (62) (147) (148). European guidelines for massive haemorrhage in severe trauma recommend FC 3-4g or Cryoprecipitate 50mg/kg to restore fibrinogen levels (64). Taneka et al,

describe in detail the rationale behind cryoprecipitate and FC dosing (149). Collins et al, have published a theoretical model of fibrinogen dosing with plasma, cryoprecipitate and FC (65). Although this model is not designed for clinical use it does highlight the significant differences in volume of product potentially required to achieve effective fibrinogen supplementation. One of the potential advantages of using FC is the standardised fibrinogen concentration per vial (+/- 1g); making dosing and assessment of dose response easier than when utilising cryoprecipitate, which has a very wide variability in fibrinogen content per unit (131). Dosing strategies for FC using VHA have been suggested in the cardiac surgical patient population (150). In traumatic haemorrhage a few animal studies have suggested potential dosing strategies but there is a paucity of quality human data (151) (152).

Published data and local institutional data suggest that 1g of Fibrinogen (FC or Cryoprecipitate) will result in an increment of between 1.5 and 2mm in the FIBTEM assay (123) (150) (153). This is in line with published data suggesting that a 1g dose of fibrinogen will result in a plasma fibrinogen increment of 0.25g/l (56) (135) (154). Due to the variability in fibrinogen concentration in cryoprecipitate, including frequent loss of units to breakage during thawing, accurate dosing is more feasible with FC. Our local data supports the published literature in equating 1g of FC to between 3 and 5 single Units of cryoprecipitate (149). The recently completed but not yet published FlinTIC trial (NCT01475344) may provide guidance on appropriate FC dosing; this study utilised a weight based FC dosing strategy in the pre-hospital environment with fibrinogen levels on arrival to ED as the primary endpoint (155).

A recent publication from the AUVA Trauma Centre demonstrated that patients treated with FC did not have higher plasma fibrinogen levels than the control group of trauma patients not treated with FC in the post trauma phase (up to day 7) (156). Suggesting that despite relatively high doses of FC there is no 'overshoot' in plasma fibrinogen levels beyond expected levels subsequent to severe injury. All patients exhibited a rise in plasma fibrinogen levels post trauma that can be attributed to increased hepatic synthesis as part of the acute phase response to severe trauma. These findings are supported by the CRYOSTAT study; where there was no excessive rise in plasma fibrinogen levels or increased risk of thromboembolic complications with fibrinogen supplementation using cryoprecipitate therapy (69).

Fibrinogen Trials in Non-Traumatic Haemorrhage:

The use of Fibrinogen Concentrate has been extensively investigated in randomised controlled trials in patients with post-partum haemorrhage and those undergoing cardiac surgery. In the recently published FIB-PPH study, women with post-partum haemorrhage (PPH) were randomised into receiving either 2g FC or placebo, after clinical suspicion of significant PPH (>1.5L) (157). There was no difference in any of the primary outcomes between the two groups. However, the data presented showed that the mean plasma fibrinogen level in both groups was > 4g/l and therefore hypofibrinogenaemia was unlikely to be contributing to bleeding. The REPLACE study; investigating FC use in cardiac surgery has recently been published (158). This study randomised patients to FC or Placebo with a 5 minute bleeding mass of >60g after separation from cardio-pulmonary bypass. The study

reported increased allogenic blood product requirements in the FC arm. This was a surprising finding which was incongruent with the results of the pilot single centre studies and a full explanation is not clear. It is postulated that the low observed bleeding rates, use of the 5 minute bleeding mass (not routinely used in clinical practice), normal range plasma fibrinogen levels and a complex treatment algorithm all contributed to the unexpected results of the trial. These studies suggests that based on purely clinical indications it is difficult to predict which patients may benefit from fibrinogen supplementation and there is no benefit to fibrinogen supplementation in patients with normal fibrinogen levels.

A single “one-off” dose of additional fibrinogen supplementation “one size fits all approach” may not be appropriate and it is possible that fibrinogen replacement may better be guided by the degree of hypofibrinogenaemia to avoid potential under and over dosing. In a recent study investigating fixed dose fibrinogen concentrate supplementation in cardiac surgery – The Zero Plasma Trial (ZEPLAST), it was shown that a fixed dose of 6g FC reduced post-operative bleeding and blood product transfusion (159). However, subsequent data analysis revealed that a reduced dose of FC would have likely yielded the same results (160). It would seem to make intuitive sense to dose fibrinogen replacement based on degree of hypofibrinogenaemia.

Fibrinogen Trials in Traumatic Haemorrhage:

There is increasing recognition and good evidence to support of the importance of fibrinogen in effective clot formation in severe traumatic haemorrhage. The utility of

early fibrinogen replacement using FC and/or cryoprecipitate is gaining popularity but at the current time is not supported by high quality evidence (161). In the last decade there has been a vast amount of literature published regarding traumatic coagulopathy and transfusion strategies. However, the quality of evidence remains low as many of the reported studies contain significant methodological and statistical flaws (162).

The heterogeneous nature of injury pattern, the complex nature of traumatic coagulopathy and the geographical variation in clinical practice makes performing studies logistically challenging (163). Although individual patient randomised controlled trials in trauma present unique challenges to investigators, they are possible to perform successfully (48) (69). Two large multi-centre RCT's in bleeding civilian trauma patients have shown that outcomes can be improved with rapid haemostatic intervention. The CRASH-2 trial demonstrated a significant survival benefit in trauma patients treated with Tranexamic Acid within 3 hours of injury (164). Although the PROPPR study showed no difference in the primary outcome measures there was reduced death from haemorrhage and more rapid haemorrhage control in the intervention group (48).

A number of randomised controlled trials in severe traumatic haemorrhage investigating a VHA guided approach with use of Factor Concentrates are currently underway. The RETIC Trial (Reversal of TIC using Coagulation Factors or Fresh Frozen Plasma, NCT01545635) utilised a VHA (ROTEM®) algorithm to guide blood product transfusion. The intervention arm received FC and/or PCC in response to predefined (ROTEM®) values; the control arm received Plasma transfusion in

response to the same predefined (ROTEM®) values. Transfusion of PRBC, Platelets and TXA was the same in both arms and followed standard institutional clinical practise. The primary outcome measure being difference in MOF rates between the two groups. This trial has been terminated early after an interim analysis (100 patients) revealed possible harm to patients randomised to the Plasma arm. The STATA Trial (Strategy of Transfusion in Trauma Patients, NCT02416817) is comparing a VHA guided approach to a standard fixed ratio MHP. The intervention arm will receive Factor Concentrate (FC and PCC) and Platelet resuscitation guided by a VHA (ROTEM®) algorithm. The control arm will receive blood product transfusion as per a 1:1:1 MHP. The trial is aiming to recruit 200 patients with a primary outcome measure of SOFA scores during first 5 days of hospital admission and is expected to complete early in 2017.

Three randomised controlled blinded trials investigating early Fibrinogen Concentrate replacement in severe traumatic haemorrhage are currently underway or have recently completed; FiiRST (Fibrinogen in the initial Resuscitation of Severe Trauma, NCT02203968), E-FIT1 (Early Fibrinogen in Trauma, ISRCTN67540073) and PRooF-iTH (Pilot Randomised trial of Fibrinogen in Trauma Haemorrhage, NCT02344069). The FiiRST trial has been completed but not yet published. This pilot feasibility study randomised bleeding trauma patients to receive either 6g FC or Placebo on admission to the ED with a primary outcome measure being proportion of patients receiving intervention within 1 hour of hospital admission. The E-FIT1 trial currently recruiting in the UK is also enrolling patients based on clinical likelihood of significant haemorrhage with patients randomised to a single dose of 6g FC or placebo (in addition to standard MHP) as soon as possible after ED admission. The primary outcome measures are feasibility of administering FC within 45 minutes of

ED admission and the proportion of patients with at least one FibC level $\geq 2\text{g/L}$ during active haemorrhage. The PRooF-iTH trial currently recruiting in Copenhagen is slightly different from FiiRSt and E-FIT1. This study is again randomising patients with traumatic haemorrhage to receiving 60-70mg/Kg FC or placebo immediately and pre-emptively on arrival in the ED. The primary outcome is change in TEG FF Maximum Amplitude at 15 minutes after intervention. These trials will certainly address significant gaps in the evidence base surrounding FC use in severe trauma and help plan future studies.

The CRYOSTAT 2 Trial is currently in the planning stages based on the results of the pilot feasibility CRYOSTAT trial. CRYOSTAT 2 will be a large, international, multi-centre trial in severely bleeding trauma patients investigating fibrinogen supplementation using empiric and early cryoprecipitate as part of a MHP, with mortality as the primary outcome measure.

Fibrinogen Concentrate vs Cryoprecipitate in Traumatic Haemorrhage:

The large doses of cryoprecipitate utilised in traumatic haemorrhage place a significant strain on local blood banks in issuing requested units in a timely manner and on national blood supply agencies in maintaining and providing adequate stocks to support ABO requirements for individual blood banks. In addition the size and population distribution of countries like Australia makes supplying and maintaining adequate stocks of 'fresh' blood products in remote locations logistically challenging. The use of a lyophilised fibrinogen factor concentrate that has a long shelf life and is

easy to use has enormous implications for both large urban metropolitan areas and remote isolated communities.

However, randomised controlled trials are urgently required to investigate the haemostatic efficacy of cryoprecipitate compared to fibrinogen concentrate in traumatic haemorrhage. A recently published systematic review found sparse evidence comparing FC to cryoprecipitate and concluded that it was not possible to recommend one product over another in bleeding associated with acquired hypofibrinogenaemia (161). It is imperative that robust and clinically relevant studies are performed before widespread practice changes are implemented without a solid evidence base that would subsequently make performing such studies unfeasible (140) (165).

Recently published negative studies investigating the blind administration of fibrinogen supplementation in severe haemorrhage combined with the delays in fibrinogen replacement utilising MHP or standard laboratory tests potentially justify the conduct of a VHA guided trial. Although the inclusion criteria of the FiiRST, E-FIT1 and PRooF-iTH trials are robust and will identify patients with significant haemorrhage, it may not be possible to identify those patients in which hypofibrinogenaemia is contributing to on-going TIC associated haemorrhage. It is imperative to ensure that the appropriate product is given at the appropriate time in the appropriate quantity (157). It is likely that only sustained fibrinogen replacement throughout the resuscitation period in tandem with surgical haemorrhage control has the potential to impact on hypofibrinogenaemia and TIC (166). The use of a directed dosing strategy could inform dose response relationships for both FC and

cryoprecipitate in terms of plasma fibrinogen increments. Additionally it has been suggested that future trauma studies could use admission FIBTEM A5 measures to dose adjust fibrinogen replacement (69).

The Fibrinogen Early In Severe Trauma study (FEISTY, NCT02745041) is a pilot, multi-centre, randomised controlled trial comparing FC to cryoprecipitate for fibrinogen supplementation in severe traumatic haemorrhage using accepted VHA triggers. This pragmatic study expanding on the currently utilised approach at the study sites; investigating the feasibility and efficacy of early fibrinogen supplementation is the first RCT comparing FC to Cryoprecipitate in traumatic haemorrhage. Adult patients with severe trauma and evidence of significant haemorrhage will be enrolled on arrival to the trauma unit and randomised to receiving fibrinogen supplementation with either FC or cryoprecipitate (Figure 5). The primary outcomes are time to administration of fibrinogen supplementation from time of ROTEM analysis (and clinical scenario) indicating fibrinogen supplementation is required and effects of fibrinogen supplementation on fibrinogen levels. Secondary outcomes include; blood product transfusion requirements, thromboembolic complications, hospital length of stay and mortality. A number of feasibility outcome measures will also be assessed. The study will take place in 4 major Queensland trauma centres and is expected to start recruiting in October 2016. The results of FEISTY will be used to design a larger and hopefully definitive multi-centre study with the aim of addressing patient centred outcomes such as allogenic blood product transfusion requirements and mortality. By performing a pilot multi-centre study it is hoped to identify logistical issues that could impact on the design and conduct of a

definitive study and potentially avoid the mistakes made in recently published FC studies (158).

Conclusion:

Whilst early fibrinogen supplementation with a concentrated product in severe traumatic haemorrhage is an attractive therapeutic option, there is currently inadequate high-level evidence to support its use. A number of on-going studies are currently investigating early fibrinogen replacement in severe trauma. Although the majority are pilot feasibility studies they will assist in planning larger definitive trials for the benefit of individual patients affected by trauma and for the community as a whole.

Funding: The programme of trauma research is supported by grants from the Emergency Medicine Foundation, National Blood Authority and Gold Coast University Hospital Foundation.

Attestation: All authors attest to having approved the final manuscript.

Conflict of Interest: Dr James Winearls has received educational, travel and research support from TEM International and CSL Behring. All other authors have no conflicts to disclose.

Acknowledgements:

We acknowledge all members of the FEISTY research team; Elizabeth Wake (GCUH), Kerin Walters (GCUH), Jody Paxton (GCUH), Mandy Tallott (GCUH), Sarah Czuchwicki (GCUH), Anthony Ghent (GCUH) and Phillip Mondy (ARCBS)

References:

1. Norton R, Kobusingye O. Injuries. *The New England journal of medicine*. 2013;368(18):1723-30.
2. Cothren CC, Moore EE, Hedegaard HB, Meng K. Epidemiology of urban trauma deaths: a comprehensive reassessment 10 years later. *World journal of surgery*. 2007;31(7):1507-11.
3. Kauvar DS, Wade CE. The epidemiology and modern management of traumatic hemorrhage: US and international perspectives. *Critical care (London, England)*. 2005;9 Suppl 5:S1-9.
4. Tien HC, Spencer F, Tremblay LN, Rizoli SB, Brenneman FD. Preventable deaths from hemorrhage at a level I Canadian trauma center. *The Journal of trauma*. 2007;62(1):142-6.
5. Minei JP, Cuschieri J, Sperry J, Moore EE, West MA, Harbrecht BG, et al. The changing pattern and implications of multiple organ failure after blunt injury with hemorrhagic shock. *Critical care medicine*. 2012;40(4):1129-35.
6. Holena DN, Netzer G, Localio R, Gallop RJ, Bellamy SL, Meyer NJ, et al. The association of early transfusion with acute lung injury in patients with severe injury. *The journal of trauma and acute care surgery*. 2012;73(4):825-31.
7. Watson GA, Sperry JL, Rosengart MR, Minei JP, Harbrecht BG, Moore EE, et al. Fresh frozen plasma is independently associated with a higher risk of multiple organ failure and acute respiratory distress syndrome. *The Journal of trauma*. 2009;67(2):221-7; discussion 8-30.
8. Brohi K, Cohen MJ, Davenport RA. Acute coagulopathy of trauma: mechanism, identification and effect. *Current opinion in critical care*. 2007;13(6):680-5.
9. Simmons JW, Pittet JF, Pierce B. Trauma-Induced Coagulopathy. *Current anesthesiology reports*. 2014;4(3):189-99.
10. Christiaans SC, Duhachek-Stapelman AL, Russell RT, Lisco SJ, Kerby JD, Pittet JF. Coagulopathy after severe pediatric trauma. *Shock (Augusta, Ga)*. 2014;41(6):476-90.
11. Dobson GP, Letson HL, Sharma R, Sheppard FR, Cap AP. Mechanisms of early trauma-induced coagulopathy: The clot thickens or not? *The journal of trauma and acute care surgery*. 2015;79(2):301-9.
12. Davenport RA, Brohi K. Cause of trauma-induced coagulopathy. *Current opinion in anaesthesiology*. 2015.
13. Brohi K, Cohen MJ, Ganter MT, Schultz MJ, Levi M, Mackersie RC, et al. Acute coagulopathy of trauma: hypoperfusion induces systemic anticoagulation and hyperfibrinolysis. *The Journal of trauma*. 2008;64(5):1211-7; discussion 7.
14. Cap A, Hunt BJ. The pathogenesis of traumatic coagulopathy. *Anaesthesia*. 2015;70 Suppl 1:96-e34.
15. Brohi K, Singh J, Heron M, Coats T. Acute traumatic coagulopathy. *The Journal of trauma*. 2003;54(6):1127-30.
16. MacLeod JB, Lynn M, McKenney MG, Cohn SM, Murtha M. Early coagulopathy predicts mortality in trauma. *The Journal of trauma*. 2003;55(1):39-44.

17. Hoffman M, Monroe DM, 3rd. A cell-based model of hemostasis. *Thrombosis and haemostasis*. 2001;85(6):958-65.
18. Lowe GD, Rumley A, Mackie IJ. Plasma fibrinogen. *Annals of clinical biochemistry*. 2004;41(Pt 6):430-40.
19. Mosesson MW. Fibrinogen and fibrin structure and functions. *Journal of thrombosis and haemostasis : JTH*. 2005;3(8):1894-904.
20. Meyer MA, Ostrowski SR, Windelov NA, Johansson PI. Fibrinogen concentrates for bleeding trauma patients: what is the evidence? *Vox sanguinis*. 2011;101(3):185-90.
21. Aubron C, Reade MC, Fraser JF, Cooper DJ. Efficacy and safety of fibrinogen concentrate in trauma patients--a systematic review. *Journal of critical care*. 2014;29(3):471 e11-7.
22. Hagemo JS, Stanworth S, Juffermans NP, Brohi K, Cohen M, Johansson PI, et al. Prevalence, predictors and outcome of hypofibrinogenaemia in trauma: a multicentre observational study. *Critical care (London, England)*. 2014;18(2):R52.
23. Hiippala ST, Myllylä GJ, Vahtera EM. Hemostatic factors and replacement of major blood loss with plasma-poor red cell concentrates. *Anesthesia and analgesia*. 1995;81(2):360-5.
24. Floccard B, Rugeri L, Faure A, Saint Denis M, Boyle EM, Peguet O, et al. Early coagulopathy in trauma patients: an on-scene and hospital admission study. *Injury*. 2012;43(1):26-32.
25. Kimura Y, Kimura S, Sumita S, Yamakage M. Predictors of hypofibrinogenemia in blunt trauma patients on admission. *Journal of anesthesia*. 2015;29(2):242-8.
26. Deras P, Villiet M, Manzanera J, Latry P, Schved JF, Capdevila X, et al. Early coagulopathy at hospital admission predicts initial or delayed fibrinogen deficit in severe trauma patients. *The journal of trauma and acute care surgery*. 2014;77(3):433-40.
27. Martini WZ, Pusateri AE, Uscilowicz JM, Delgado AV, Holcomb JB. Independent contributions of hypothermia and acidosis to coagulopathy in swine. *The Journal of trauma*. 2005;58(5):1002-9; discussion 9-10.
28. Fenger-Eriksen C, Tonnesen E, Ingerslev J, Sorensen B. Mechanisms of hydroxyethyl starch-induced dilutional coagulopathy. *Journal of thrombosis and haemostasis : JTH*. 2009;7(7):1099-105.
29. Martini WZ, Holcomb JB. Acidosis and coagulopathy: the differential effects on fibrinogen synthesis and breakdown in pigs. *Annals of surgery*. 2007;246(5):831-5.
30. Martini WZ. The effects of hypothermia on fibrinogen metabolism and coagulation function in swine. *Metabolism: clinical and experimental*. 2007;56(2):214-21.
31. Kashuk JL, Moore EE, Sawyer M, Wohlauser M, Pezold M, Barnett C, et al. Primary fibrinolysis is integral in the pathogenesis of the acute coagulopathy of trauma. *Annals of surgery*. 2010;252(3):434-42; discussion 43-4.
32. Levrat A, Gros A, Rugeri L, Inaba K, Floccard B, Negrier C, et al. Evaluation of rotation thrombelastography for the diagnosis of hyperfibrinolysis in trauma patients. *British journal of anaesthesia*. 2008;100(6):792-7.
33. Theusinger OM, Wanner GA, Emmert MY, Billeter A, Eismann J, Seifert B, et al. Hyperfibrinolysis diagnosed by rotational thromboelastometry (ROTEM) is associated with higher mortality in patients with severe trauma. *Anesthesia and analgesia*. 2011;113(5):1003-12.
34. Cotton BA, Harvin JA, Kostousou V, Minei KM, Radwan ZA, Schochl H, et al. Hyperfibrinolysis at admission is an uncommon but highly lethal event associated with shock and prehospital fluid administration. *The journal of trauma and acute care surgery*. 2012;73(2):365-70; discussion 70.
35. Raza I, Davenport R, Rourke C, Platton S, Manson J, Spoors C, et al. The incidence and magnitude of fibrinolytic activation in trauma patients. *Journal of thrombosis and haemostasis : JTH*. 2013;11(2):307-14.
36. Schochl H, Frietsch T, Pavelka M, Jambor C. Hyperfibrinolysis after major trauma: differential diagnosis of lysis patterns and prognostic value of thrombelastometry. *The Journal of trauma*. 2009;67(1):125-31.

37. Rourke C, Curry N, Khan S, Taylor R, Raza I, Davenport R, et al. Fibrinogen levels during trauma hemorrhage, response to replacement therapy, and association with patient outcomes. *Journal of thrombosis and haemostasis : JTH*. 2012;10(7):1342-51.
38. Inaba K, Karamanos E, Lustenberger T, Schochl H, Shulman I, Nelson J, et al. Impact of fibrinogen levels on outcomes after acute injury in patients requiring a massive transfusion. *Journal of the American College of Surgeons*. 2013;216(2):290-7.
39. Holcomb JB, Jenkins D, Rhee P, Johannigman J, Mahoney P, Mehta S, et al. Damage control resuscitation: directly addressing the early coagulopathy of trauma. *The Journal of trauma*. 2007;62(2):307-10.
40. Holcomb JB. Optimal use of blood products in severely injured trauma patients. *Hematology / the Education Program of the American Society of Hematology American Society of Hematology Education Program*. 2010;2010:465-9.
41. Gonzalez EA, Moore FA, Holcomb JB, Miller CC, Kozar RA, Todd SR, et al. Fresh frozen plasma should be given earlier to patients requiring massive transfusion. *The Journal of trauma*. 2007;62(1):112-9.
42. Borgman MA, Spinella PC, Perkins JG, Grathwohl KW, Repine T, Beekley AC, et al. The ratio of blood products transfused affects mortality in patients receiving massive transfusions at a combat support hospital. *The Journal of trauma*. 2007;63(4):805-13.
43. Holcomb JB, Wade CE, Michalek JE, Chisholm GB, Zarzabal LA, Schreiber MA, et al. Increased plasma and platelet to red blood cell ratios improves outcome in 466 massively transfused civilian trauma patients. *Annals of surgery*. 2008;248(3):447-58.
44. Snyder CW, Weinberg JA, McGwin G, Jr., Melton SM, George RL, Reiff DA, et al. The relationship of blood product ratio to mortality: survival benefit or survival bias? *The Journal of trauma*. 2009;66(2):358-62; discussion 62-4.
45. Schuster KM, Davis KA, Lui FY, Maerz LL, Kaplan LJ. The status of massive transfusion protocols in United States trauma centers: massive transfusion or massive confusion? *Transfusion*. 2010;50(7):1545-51.
46. Callum JL, Nascimento B, Alam A. Massive haemorrhage protocol: what's the best protocol? *ISBT Science Series*. 2016;11:297-306.
47. Holcomb JB, del Junco DJ, Fox EE, Wade CE, Cohen MJ, Schreiber MA, et al. The prospective, observational, multicenter, major trauma transfusion (PROMMTT) study: comparative effectiveness of a time-varying treatment with competing risks. *JAMA surgery*. 2013;148(2):127-36.
48. Holcomb JB, Tilley BC, Baraniuk S, Fox EE, Wade CE, Podbielski JM, et al. Transfusion of plasma, platelets, and red blood cells in a 1:1:1 vs a 1:1:2 ratio and mortality in patients with severe trauma: the PROPPR randomized clinical trial. *Jama*. 2015;313(5):471-82.
49. Dirks J, Jorgensen H, Jensen CH, Ostrowski SR, Johansson PI. Blood product ratio in acute traumatic coagulopathy--effect on mortality in a Scandinavian level 1 trauma centre. *Scandinavian journal of trauma, resuscitation and emergency medicine*. 2010;18:65.
50. Davenport R, Curry N, Manson J, De'Ath H, Coates A, Rourke C, et al. Hemostatic effects of fresh frozen plasma may be maximal at red cell ratios of 1:2. *The Journal of trauma*. 2011;70(1):90-5; discussion 5-6.
51. Inaba K, Branco BC, Rhee P, Blackburne LH, Holcomb JB, Teixeira PG, et al. Impact of plasma transfusion in trauma patients who do not require massive transfusion. *Journal of the American College of Surgeons*. 2010;210(6):957-65.
52. Novak DJ, Bai Y, Cooke RK, Marques MB, Fontaine MJ, Gottschall JL, et al. Making thawed universal donor plasma available rapidly for massively bleeding trauma patients: experience from the Pragmatic, Randomized Optimal Platelets and Plasma Ratios (PROPPR) trial. *Transfusion*. 2015;55(6):1331-9.
53. Nascimento B, Callum J, Tien H, Rubenfeld G, Pinto R, Lin Y, et al. Effect of a fixed-ratio (1:1:1) transfusion protocol versus laboratory-results-guided transfusion in patients with severe

trauma: a randomized feasibility trial. *CMAJ : Canadian Medical Association journal = journal de l'Association medicale canadienne*. 2013;185(12):E583-9.

54. Stanworth SJ, Davenport R, Curry N, Seeney F, Eaglestone S, Edwards A, et al. Mortality from trauma haemorrhage and opportunities for improvement in transfusion practice. *The British journal of surgery*. 2016;103(4):357-65.
55. Schochl H, Cotton B, Inaba K, Nienaber U, Fischer H, Voelckel W, et al. FIBTEM provides early prediction of massive transfusion in trauma. *Critical care (London, England)*. 2011;15(6):R265.
56. Fenger-Eriksen C, Lindberg-Larsen M, Christensen AQ, Ingerslev J, Sorensen B. Fibrinogen concentrate substitution therapy in patients with massive haemorrhage and low plasma fibrinogen concentrations. *British journal of anaesthesia*. 2008;101(6):769-73.
57. Innerhofer P, Westermann I, Tauber H, Breitenkopf R, Fries D, Kastenberger T, et al. The exclusive use of coagulation factor concentrates enables reversal of coagulopathy and decreases transfusion rates in patients with major blunt trauma. *Injury*. 2013;44(2):209-16.
58. Schochl H, Forster L, Woidke R, Solomon C, Voelckel W. Use of rotation thromboelastometry (ROTEM) to achieve successful treatment of polytrauma with fibrinogen concentrate and prothrombin complex concentrate. *Anaesthesia*. 2010;65(2):199-203.
59. Stinger HK, Spinella PC, Perkins JG, Grathwohl KW, Salinas J, Martini WZ, et al. The ratio of fibrinogen to red cells transfused affects survival in casualties receiving massive transfusions at an army combat support hospital. *The Journal of trauma*. 2008;64(2 Suppl):S79-85; discussion S.
60. Morrison JJ, Ross JD, Dubose JJ, Jansen JO, Midwinter MJ, Rasmussen TE. Association of cryoprecipitate and tranexamic acid with improved survival following wartime injury: findings from the MATTERS II Study. *JAMA surgery*. 2013;148(3):218-25.
61. Chowdhary P, Saayman AG, Paulus U, Findlay GP, Collins PW. Efficacy of standard dose and 30 ml/kg fresh frozen plasma in correcting laboratory parameters of haemostasis in critically ill patients. *British journal of haematology*. 2004;125(1):69-73.
62. Nascimento B, Goodnough LT, Levy JH. Cryoprecipitate therapy. *British journal of anaesthesia*. 2014;113(6):922-34.
63. Levy JH, Welsby I, Goodnough LT. Fibrinogen as a therapeutic target for bleeding: a review of critical levels and replacement therapy. *Transfusion*. 2014;54(5):1389-405; quiz 8.
64. Rossaint R, Bouillon B, Cerny V, Coats TJ, Duranteau J, Fernandez-Mondejar E, et al. The European guideline on management of major bleeding and coagulopathy following trauma: fourth edition. *Critical care (London, England)*. 2016;20(1):100.
65. Collins PW, Solomon C, Sutor K, Crispin D, Hochleitner G, Rizoli S, et al. Theoretical modelling of fibrinogen supplementation with therapeutic plasma, cryoprecipitate, or fibrinogen concentrate. *British journal of anaesthesia*. 2014;113(4):585-95.
66. Khan S, Davenport R, Raza I, Glasgow S, De'Ath HD, Johansson PI, et al. Damage control resuscitation using blood component therapy in standard doses has a limited effect on coagulopathy during trauma hemorrhage. *Intensive care medicine*. 2015;41(2):239-47.
67. Chambers LA, Chow SJ, Shaffer LE. Frequency and characteristics of coagulopathy in trauma patients treated with a low- or high-plasma-content massive transfusion protocol. *American journal of clinical pathology*. 2011;136(3):364-70.
68. Holcomb JB, Fox EE, Zhang X, White N, Wade CE, Cotton BA, et al. Cryoprecipitate use in the PROMMTT study. *The journal of trauma and acute care surgery*. 2013;75(1 Suppl 1):S31-9.
69. Curry N, Rourke C, Davenport R, Beer S, Pankhurst L, Deary A, et al. Early cryoprecipitate for major haemorrhage in trauma: a randomised controlled feasibility trial. *British journal of anaesthesia*. 2015;115(1):76-83.
70. Haas T, Fries D, Tanaka KA, Asmis L, Curry NS, Schochl H. Usefulness of standard plasma coagulation tests in the management of perioperative coagulopathic bleeding: is there any evidence? *British journal of anaesthesia*. 2015;114(2):217-24.

71. Toulon P, Ozier Y, Ankri A, Fleron MH, Leroux G, Samama CM. Point-of-care versus central laboratory coagulation testing during haemorrhagic surgery. A multicenter study. *Thrombosis and haemostasis*. 2009;101(2):394-401.
72. Adam S, Karger R, Kretschmer V. Photo-optical methods can lead to clinically relevant overestimation of fibrinogen concentration in plasma diluted with hydroxyethyl starch. *Clinical and applied thrombosis/hemostasis : official journal of the International Academy of Clinical and Applied Thrombosis/Hemostasis*. 2010;16(4):461-71.
73. Schlomp CJ, Schochl H. The role of fibrinogen in trauma-induced coagulopathy. *Hamostaseologie*. 2014;34(1):29-39.
74. Mackie IJ, Kitchen S, Machin SJ, Lowe GD. Guidelines on fibrinogen assays. *British journal of haematology*. 2003;121(3):396-404.
75. Solomon C, Baryshnikova E, Tripodi A, Schlomp CJ, Schochl H, Cadamuro J, et al. Fibrinogen measurement in cardiac surgery with cardiopulmonary bypass: analysis of repeatability and agreement of Clauss method within and between six different laboratories. *Thrombosis and haemostasis*. 2014;112(1):109-17.
76. Hiippala ST. Dextran and hydroxyethyl starch interfere with fibrinogen assays. *Blood coagulation & fibrinolysis : an international journal in haemostasis and thrombosis*. 1995;6(8):743-6.
77. Fenger-Eriksen C, Moore GW, Rangarajan S, Ingerslev J, Sorensen B. Fibrinogen estimates are influenced by methods of measurement and hemodilution with colloid plasma expanders. *Transfusion*. 2010;50(12):2571-6.
78. Davenport R, Manson J, De'Ath H, Platton S, Coates A, Allard S, et al. Functional definition and characterization of acute traumatic coagulopathy. *Critical care medicine*. 2011;39(12):2652-8.
79. Chandler WL, Ferrell C, Trimble S, Moody S. Development of a rapid emergency hemorrhage panel. *Transfusion*. 2010;50(12):2547-52.
80. Johansson PI, Stensballe J. Effect of Haemostatic Control Resuscitation on mortality in massively bleeding patients: a before and after study. *Vox sanguinis*. 2009;96(2):111-8.
81. Inaba K, Rizoli S, Veigas PV, Callum J, Davenport R, Hess J, et al. 2014 Consensus conference on viscoelastic test-based transfusion guidelines for early trauma resuscitation: Report of the panel. *The journal of trauma and acute care surgery*. 2015;78(6):1220-9.
82. Weber CF, Gorlinger K, Meininger D, Herrmann E, Bingold T, Moritz A, et al. Point-of-care testing: a prospective, randomized clinical trial of efficacy in coagulopathic cardiac surgery patients. *Anesthesiology*. 2012;117(3):531-47.
83. Schaden E, Kimberger O, Kraincuk P, Baron DM, Metnitz PG, Kozek-Langenecker S. Perioperative treatment algorithm for bleeding burn patients reduces allogeneic blood product requirements. *British journal of anaesthesia*. 2012;109(3):376-81.
84. Winearls J, Reade M, Miles H, Bulmer A, Campbell D, Gorlinger K, et al. Targeted Coagulation Management in Severe Trauma: The Controversies and the Evidence. *Anesthesia and analgesia*. 2016;123(4):910-24.
85. Tanaka KA, Bolliger D, Vadlamudi R, Nimmo A. Rotational thromboelastometry (ROTEM)-based coagulation management in cardiac surgery and major trauma. *Journal of cardiothoracic and vascular anesthesia*. 2012;26(6):1083-93.
86. Abdelfattah K, Cripps MW. Thromboelastography and Rotational Thromboelastometry use in trauma. *International journal of surgery (London, England)*. 2015.
87. Afshari A, Wikkelsø A, Brok J, Møller AM, Wetterslev J. Thromboelastography (TEG) or thromboelastometry (ROTEM) to monitor haemotherapy versus usual care in patients with massive transfusion. *The Cochrane database of systematic reviews*. 2011(3):CD007871.
88. Wolberg AS, Campbell RA. Thrombin generation, fibrin clot formation and hemostasis. *Transfusion and apheresis science : official journal of the World Apheresis Association : official journal of the European Society for Haemapheresis*. 2008;38(1):15-23.

89. Rizoli S, Min A, Sanchez AP, Shek P, Grodecki R, Veigas P, et al. In Trauma, Conventional ROTEM and TEG Results Are Not Interchangeable But Are Similar in Clinical Applicability. *Military medicine*. 2016;181(5 Suppl):117-26.
90. Hagemo JS, Christiaans SC, Stanworth SJ, Brohi K, Johansson PI, Goslings JC, et al. Detection of acute traumatic coagulopathy and massive transfusion requirements by means of rotational thromboelastometry: an international prospective validation study. *Critical care (London, England)*. 2015;19:97.
91. Johansson PI, Stissing T, Bochsén L, Ostrowski SR. Thrombelastography and thromboelastometry in assessing coagulopathy in trauma. *Scandinavian journal of trauma, resuscitation and emergency medicine*. 2009;17:45.
92. Tauber H, Innerhofer P, Breitkopf R, Westermann I, Beer R, El Attal R, et al. Prevalence and impact of abnormal ROTEM(R) assays in severe blunt trauma: results of the 'Diagnosis and Treatment of Trauma-Induced Coagulopathy (DIA-TRE-TIC) study'. *British journal of anaesthesia*. 2011;107(3):378-87.
93. Holcomb JB, Minei KM, Scerbo ML, Radwan ZA, Wade CE, Kozar RA, et al. Admission rapid thrombelastography can replace conventional coagulation tests in the emergency department: experience with 1974 consecutive trauma patients. *Annals of surgery*. 2012;256(3):476-86.
94. Goodman MD, Makley AT, Hanseman DJ, Pritts TA, Robinson BR. All the bang without the bucks: Defining essential point-of-care testing for traumatic coagulopathy. *The journal of trauma and acute care surgery*. 2015;79(1):117-24; discussion 24.
95. Schochl H, Maegele M, Solomon C, Gorlinger K, Voelckel W. Early and individualized goal-directed therapy for trauma-induced coagulopathy. *Scandinavian journal of trauma, resuscitation and emergency medicine*. 2012;20:15.
96. Schochl H, Nienaber U, Hofer G, Voelckel W, Jambor C, Scharbert G, et al. Goal-directed coagulation management of major trauma patients using thromboelastometry (ROTEM)-guided administration of fibrinogen concentrate and prothrombin complex concentrate. *Critical care (London, England)*. 2010;14(2):R55.
97. Gonzalez E, Moore EE, Moore HB, Chapman MP, Chin TL, Ghasabyan A, et al. Goal-directed Hemostatic Resuscitation of Trauma-induced Coagulopathy: A Pragmatic Randomized Clinical Trial Comparing a Viscoelastic Assay to Conventional Coagulation Assays. *Annals of surgery*. 2015.
98. Schochl H, Nienaber U, Maegele M, Hochleitner G, Primavesi F, Steitz B, et al. Transfusion in trauma: thromboelastometry-guided coagulation factor concentrate-based therapy versus standard fresh frozen plasma-based therapy. *Critical care (London, England)*. 2011;15(2):R83.
99. Schlump CJ, Voelckel W, Inaba K, Maegele M, Schochl H. Impact of fibrinogen concentrate alone or with prothrombin complex concentrate (+/- fresh frozen plasma) on plasma fibrinogen level and fibrin-based clot strength (FIBTEM) in major trauma: a retrospective study. *Scandinavian journal of trauma, resuscitation and emergency medicine*. 2013;21:74.
100. Da Luz LT, Nascimento B, Shankarakutty AK, Rizoli S, Adhikari NK. Effect of thromboelastography (TEG(R)) and rotational thromboelastometry (ROTEM(R)) on diagnosis of coagulopathy, transfusion guidance and mortality in trauma: descriptive systematic review. *Critical care (London, England)*. 2014;18(5):518.
101. Wikkelsø A, Wetterslev J, Møller AM, Afshari A. Thromboelastography (TEG) or thromboelastometry (ROTEM) to monitor haemostatic treatment versus usual care in adults or children with bleeding. *The Cochrane database of systematic reviews*. 2016(8):Cd007871.
102. Hunt H, Stanworth S, Curry N, Woolley T, Cooper C, Ukoumunne O, et al. Thromboelastography (TEG) and rotational thromboelastometry (ROTEM) for trauma induced coagulopathy in adult trauma patients with bleeding. *The Cochrane database of systematic reviews*. 2015;2:CD010438.
103. Kitchen DP, Kitchen S, Jennings I, Woods T, Walker I. Quality assurance and quality control of thrombelastography and rotational Thromboelastometry: the UK NEQAS for blood coagulation experience. *Seminars in thrombosis and hemostasis*. 2010;36(7):757-63.

104. Haas T, Spielmann N, Mauch J, Speer O, Schmugge M, Weiss M. Reproducibility of thrombelastometry (ROTEM(R)): point-of-care versus hospital laboratory performance. *Scandinavian journal of clinical and laboratory investigation*. 2012;72(4):313-7.
105. Theusinger OM, Nurnberg J, Asmis LM, Seifert B, Spahn DR. Rotation thromboelastometry (ROTEM) stability and reproducibility over time. *European journal of cardio-thoracic surgery : official journal of the European Association for Cardio-thoracic Surgery*. 2010;37(3):677-83.
106. Mauch J, Spielmann N, Hartnack S, Madjdpour C, Kutter AP, Bettschart-Wolfensberger R, et al. Intrarater and interrater variability of point of care coagulation testing using the ROTEM delta. *Blood coagulation & fibrinolysis : an international journal in haemostasis and thrombosis*. 2011;22(8):662-6.
107. Meyer AS, Meyer MA, Sorensen AM, Rasmussen LS, Hansen MB, Holcomb JB, et al. Thrombelastography and rotational thromboelastometry early amplitudes in 182 trauma patients with clinical suspicion of severe injury. *The journal of trauma and acute care surgery*. 2014;76(3):682-90.
108. Spahn DR, Bouillon B, Cerny V, Coats TJ, Duranteau J, Fernandez-Mondejar E, et al. Management of bleeding and coagulopathy following major trauma: an updated European guideline. *Critical care (London, England)*. 2013;17(2):R76.
109. Rugeri L, Levrat A, David JS, Delecroix E, Floccard B, Gros A, et al. Diagnosis of early coagulation abnormalities in trauma patients by rotation thrombelastography. *Journal of thrombosis and haemostasis : JTH*. 2007;5(2):289-95.
110. Ogawa S, Szlam F, Chen EP, Nishimura T, Kim H, Roback JD, et al. A comparative evaluation of rotation thromboelastometry and standard coagulation tests in hemodilution-induced coagulation changes after cardiac surgery. *Transfusion*. 2012;52(1):14-22.
111. Rouillet S, Pillot J, Freyburger G, Biais M, Quinart A, Rault A, et al. Rotation thromboelastometry detects thrombocytopenia and hypofibrinogenaemia during orthotopic liver transplantation. *British journal of anaesthesia*. 2010;104(4):422-8.
112. Ogawa S, Szlam F, Bolliger D, Nishimura T, Chen EP, Tanaka KA. The impact of hematocrit on fibrin clot formation assessed by rotational thromboelastometry. *Anesthesia and analgesia*. 2012;115(1):16-21.
113. Meyer MA, Ostrowski SR, Sorensen AM, Meyer AS, Holcomb JB, Wade CE, et al. Fibrinogen in trauma, an evaluation of thrombelastography and rotational thromboelastometry fibrinogen assays. *The Journal of surgical research*. 2015;194(2):581-90.
114. Gorlinger K, Dirkmann D, Hanke AA, Kamler M, Kottenberg E, Thielmann M, et al. First-line therapy with coagulation factor concentrates combined with point-of-care coagulation testing is associated with decreased allogeneic blood transfusion in cardiovascular surgery: a retrospective, single-center cohort study. *Anesthesiology*. 2011;115(6):1179-91.
115. Girdauskas E, Kempfert J, Kuntze T, Borger MA, Enders J, Fassl J, et al. Thromboelastometrically guided transfusion protocol during aortic surgery with circulatory arrest: a prospective, randomized trial. *The Journal of thoracic and cardiovascular surgery*. 2010;140(5):1117-24 e2.
116. Noval-Padillo JA, Leon-Justel A, Mellado-Miras P, Porras-Lopez F, Villegas-Duque D, Gomez-Bravo MA, et al. Introduction of fibrinogen in the treatment of hemostatic disorders during orthotopic liver transplantation: implications in the use of allogenic blood. *Transplantation proceedings*. 2010;42(8):2973-4.
117. Rahe-Meyer N, Pichlmaier M, Haverich A, Solomon C, Winterhalter M, Piepenbrock S, et al. Bleeding management with fibrinogen concentrate targeting a high-normal plasma fibrinogen level: a pilot study. *British journal of anaesthesia*. 2009;102(6):785-92.
118. Gorlinger K, Dirkmann D, Solomon C, Hanke AA. Fast interpretation of thromboelastometry in non-cardiac surgery: reliability in patients with hypo-, normo-, and hypercoagulability. *British journal of anaesthesia*. 2013;110(2):222-30.

119. Pool JG, Gershgold EJ, Pappenhagen AR. HIGH-POTENCY ANTIHAEMOPHILIC FACTOR CONCENTRATE PREPARED FROM CRYOGLOBULIN PRECIPITATE. *Nature*. 1964;203:312.
120. Levy JH, Szlam F, Tanaka KA, Sniecinski RM. Fibrinogen and hemostasis: a primary hemostatic target for the management of acquired bleeding. *Anesthesia and analgesia*. 2012;114(2):261-74.
121. Tinegate H, Allard S, Grant-Casey J, Hennem S, Kilner M, Rowley M, et al. Cryoprecipitate for transfusion: which patients receive it and why? A study of patterns of use across three regions in England. *Transfusion medicine (Oxford, England)*. 2012;22(5):356-61.
122. Pantanowitz L, Kruskall MS, Uhl L. Cryoprecipitate. Patterns of use. *American journal of clinical pathology*. 2003;119(6):874-81.
123. Lee SH, Lee SM, Kim CS, Cho HS, Lee JH, Lee CH, et al. Fibrinogen recovery and changes in fibrin-based clot firmness after cryoprecipitate administration in patients undergoing aortic surgery involving deep hypothermic circulatory arrest. *Transfusion*. 2014;54(5):1379-87.
124. Levi M, Toh CH, Thachil J, Watson HG. Guidelines for the diagnosis and management of disseminated intravascular coagulation. British Committee for Standards in Haematology. *British journal of haematology*. 2009;145(1):24-33.
125. Nascimento B, Rizoli S, Rubenfeld G, Fukushima R, Ahmed N, Nathens A, et al. Cryoprecipitate transfusion: assessing appropriateness and dosing in trauma. *Transfusion medicine (Oxford, England)*. 2011;21(6):394-401.
126. Alport EC, Callum JL, Nahirniak S, Eurich B, Hume HA. Cryoprecipitate use in 25 Canadian hospitals: commonly used outside of the published guidelines. *Transfusion*. 2008;48(10):2122-7.
127. Sorensen B, Bevan D. A critical evaluation of cryoprecipitate for replacement of fibrinogen. *British journal of haematology*. 2010;149(6):834-43.
128. Ranucci M, Solomon C. Supplementation of fibrinogen in acquired bleeding disorders: experience, evidence, guidelines, and licences. *British journal of anaesthesia*. 2012;109(2):135-7.
129. Callum JL, Karkouti K, Lin Y. Cryoprecipitate: the current state of knowledge. *Transfusion medicine reviews*. 2009;23(3):177-88.
130. Faraday N. Fibrinogen concentrate and allogeneic blood transfusion in high-risk surgery. *Anesthesiology*. 2013;118(1):7-9.
131. Theodoulou A, Berryman J, Nathwani A, Scully M. Comparison of cryoprecipitate with fibrinogen concentrate for acquired hypofibrinogenaemia. *Transfusion and apheresis science : official journal of the World Apheresis Association : official journal of the European Society for Haemapheresis*. 2012;46(2):159-62.
132. Okerberg CK, Williams LA, 3rd, Kilgore ML, Kim CH, Marques MB, Schwartz J, et al. Cryoprecipitate AHF vs. fibrinogen concentrates for fibrinogen replacement in acquired bleeding patients - an economic evaluation. *Vox sanguinis*. 2016.
133. Solomon C, Hagl C, Rahe-Meyer N. Time course of haemostatic effects of fibrinogen concentrate administration in aortic surgery. *British journal of anaesthesia*. 2013;110(6):947-56.
134. Rahe-Meyer N, Solomon C, Hanke A, Schmidt DS, Knoerzer D, Hochleitner G, et al. Effects of fibrinogen concentrate as first-line therapy during major aortic replacement surgery: a randomized, placebo-controlled trial. *Anesthesiology*. 2013;118(1):40-50.
135. Fenger-Eriksen C, Jensen TM, Kristensen BS, Jensen KM, Tonnesen E, Ingerslev J, et al. Fibrinogen substitution improves whole blood clot firmness after dilution with hydroxyethyl starch in bleeding patients undergoing radical cystectomy: a randomized, placebo-controlled clinical trial. *Journal of thrombosis and haemostasis : JTH*. 2009;7(5):795-802.
136. Charbit B, Mandelbrot L, Samain E, Baron G, Haddaoui B, Keita H, et al. The decrease of fibrinogen is an early predictor of the severity of postpartum hemorrhage. *Journal of thrombosis and haemostasis : JTH*. 2007;5(2):266-73.
137. Rahe-Meyer N, Hanke A, Schmidt DS, Hagl C, Pichlmaier M. Fibrinogen concentrate reduces intraoperative bleeding when used as first-line hemostatic therapy during major aortic replacement

- surgery: results from a randomized, placebo-controlled trial. *The Journal of thoracic and cardiovascular surgery*. 2013;145(3 Suppl):S178-85.
138. Curry N, Hopewell S, Doree C, Hyde C, Brohi K, Stanworth S. The acute management of trauma hemorrhage: a systematic review of randomized controlled trials. *Critical care (London, England)*. 2011;15(2):R92.
 139. Kozek-Langenecker S, Sorensen B, Hess JR, Spahn DR. Clinical effectiveness of fresh frozen plasma compared with fibrinogen concentrate: a systematic review. *Critical care (London, England)*. 2011;15(5):R239.
 140. Wikkelsø A, Lunde J, Johansen M, Stensballe J, Wetterslev J, Møller AM, et al. Fibrinogen concentrate in bleeding patients. *The Cochrane database of systematic reviews*. 2013;8:CD008864.
 141. Nienaber U, Innerhofer P, Westermann I, Schochl H, Attal R, Breitenkopf R, et al. The impact of fresh frozen plasma vs coagulation factor concentrates on morbidity and mortality in trauma-associated haemorrhage and massive transfusion. *Injury*. 2011;42(7):697-701.
 142. Beyerle A, Nolte MW, Solomon C, Herzog E, Dickneite G. Analysis of the safety and pharmacodynamics of human fibrinogen concentrate in animals. *Toxicology and applied pharmacology*. 2014;280(1):70-7.
 143. Zentai C, Braunschweig T, Schnabel J, Rose M, Rossaint R, Grottko O. Fibrinogen concentrate does not suppress endogenous fibrinogen synthesis in a 24-hour porcine trauma model. *Anesthesiology*. 2014;121(4):753-64.
 144. Dickneite G, Pragst I, Joch C, Bergman GE. Animal model and clinical evidence indicating low thrombogenic potential of fibrinogen concentrate (Haemocomplettan P). *Blood coagulation & fibrinolysis : an international journal in haemostasis and thrombosis*. 2009;20(7):535-40.
 145. Solomon C, Groner A, Ye J, Pendrak I. Safety of fibrinogen concentrate: analysis of more than 27 years of pharmacovigilance data. *Thrombosis and haemostasis*. 2015;113(4):759-71.
 146. Lin DM, Murphy LS, Tran MH. Use of prothrombin complex concentrates and fibrinogen concentrates in the perioperative setting: a systematic review. *Transfusion medicine reviews*. 2013;27(2):91-104.
 147. Dzik WH, Blajchman MA, Fergusson D, Hameed M, Henry B, Kirkpatrick AW, et al. Clinical review: Canadian National Advisory Committee on Blood and Blood Products--Massive transfusion consensus conference 2011: report of the panel. *Critical care (London, England)*. 2011;15(6):242.
 148. Practice guidelines for perioperative blood management: an updated report by the american society of anesthesiologists task force on perioperative blood management*. *Anesthesiology*. 2015;122(2):241-75.
 149. Tanaka KA, Bader SO, Gorlinger K. Novel approaches in management of perioperative coagulopathy. *Current opinion in anaesthesiology*. 2014;27(1):72-80.
 150. Gorlinger K, Fries D, Dirkmann D, Weber CF, Hanke AA, Schochl H. Reduction of Fresh Frozen Plasma Requirements by Perioperative Point-of-Care Coagulation Management with Early Calculated Goal-Directed Therapy. *Transfusion medicine and hemotherapy : offizielles Organ der Deutschen Gesellschaft für Transfusionsmedizin und Immunhamatologie*. 2012;39(2):104-13.
 151. Martini J, Maisch S, Pilshofer L, Streif W, Martini W, Fries D. Fibrinogen concentrate in dilutional coagulopathy: a dose study in pigs. *Transfusion*. 2014;54(1):149-57.
 152. Grottko O, Braunschweig T, Henzler D, Coburn M, Tolba R, Rossaint R. Effects of different fibrinogen concentrations on blood loss and coagulation parameters in a pig model of coagulopathy with blunt liver injury. *Critical care (London, England)*. 2010;14(2):R62.
 153. Solomon C, Pichlmaier U, Schoechl H, Hagl C, Raymondos K, Scheinichen D, et al. Recovery of fibrinogen after administration of fibrinogen concentrate to patients with severe bleeding after cardiopulmonary bypass surgery. *British journal of anaesthesia*. 2010;104(5):555-62.
 154. Danes AF, Cuenca LG, Bueno SR, Mendarte Barrenechea L, Ronsano JB. Efficacy and tolerability of human fibrinogen concentrate administration to patients with acquired fibrinogen deficiency and active or in high-risk severe bleeding. *Vox sanguinis*. 2008;94(3):221-6.

155. Maegele M, Zinser M, Schlimp C, Schochl H, Fries D. Injectable hemostatic adjuncts in trauma: Fibrinogen and the FlinTIC study. *The journal of trauma and acute care surgery*. 2015;78(6 Suppl 1):S76-82.
156. Schlimp CJ, Ponschab M, Voelckel W, Treichl B, Maegele M, Schochl H. Fibrinogen levels in trauma patients during the first seven days after fibrinogen concentrate therapy: a retrospective study. *Scandinavian journal of trauma, resuscitation and emergency medicine*. 2016;24(1):29.
157. Wikkelsø AJ, Edwards HM, Afshari A, Stensballe J, Langhoff-Roos J, Albrechtsen C, et al. Pre-emptive treatment with fibrinogen concentrate for postpartum haemorrhage: randomized controlled trial. *British journal of anaesthesia*. 2015;114(4):623-33.
158. Rahe-Meyer N, Levy JH, Mazer CD, Schramko A, Klein AA, Brat R, et al. Randomized evaluation of fibrinogen vs placebo in complex cardiovascular surgery (REPLACE): a double-blind phase III study of haemostatic therapy. *British journal of anaesthesia*. 2016;117(1):41-51.
159. Ranucci M, Baryshnikova E, Crapelli GB, Rahe-Meyer N, Menicanti L, Frigiola A. Randomized, double-blinded, placebo-controlled trial of fibrinogen concentrate supplementation after complex cardiac surgery. *Journal of the American Heart Association*. 2015;4(6).
160. Ranucci M, Baryshnikova E. Fibrinogen supplementation after cardiac surgery: insights from the Zero-Plasma trial (ZEPLAST). *British journal of anaesthesia*. 2016.
161. Jensen NH, Stensballe J, Afshari A. Comparing efficacy and safety of fibrinogen concentrate to cryoprecipitate in bleeding patients: a systematic review. *Acta anaesthesiologica Scandinavica*. 2016.
162. Poole D. Coagulopathy and transfusion strategies in trauma. Overwhelmed by literature, supported by weak evidence. *Blood transfusion = Trasfusione del sangue*. 2016;14(1):3-7.
163. Schafer N, Driessen A, Frohlich M, Sturmer EK, Maegele M. Diversity in clinical management and protocols for the treatment of major bleeding trauma patients across European level I Trauma Centres. *Scandinavian journal of trauma, resuscitation and emergency medicine*. 2015;23:74.
164. Shakur H, Roberts I, Bautista R, Caballero J, Coats T, Dewan Y, et al. Effects of tranexamic acid on death, vascular occlusive events, and blood transfusion in trauma patients with significant haemorrhage (CRASH-2): a randomised, placebo-controlled trial. *Lancet*. 2010;376(9734):23-32.
165. Stanworth SJ, Hunt BJ. The desperate need for good-quality clinical trials to evaluate the optimal source and dose of fibrinogen in managing bleeding. *Critical care (London, England)*. 2011;15(6):1006.
166. Khan S, Brohi K, Chana M, Raza I, Stanworth S, Gaarder C, et al. Hemostatic resuscitation is neither hemostatic nor resuscitative in trauma hemorrhage. *The journal of trauma and acute care surgery*. 2014;76(3):561-7; discussion 7-8.

Figure 1. ROTEM Temogram

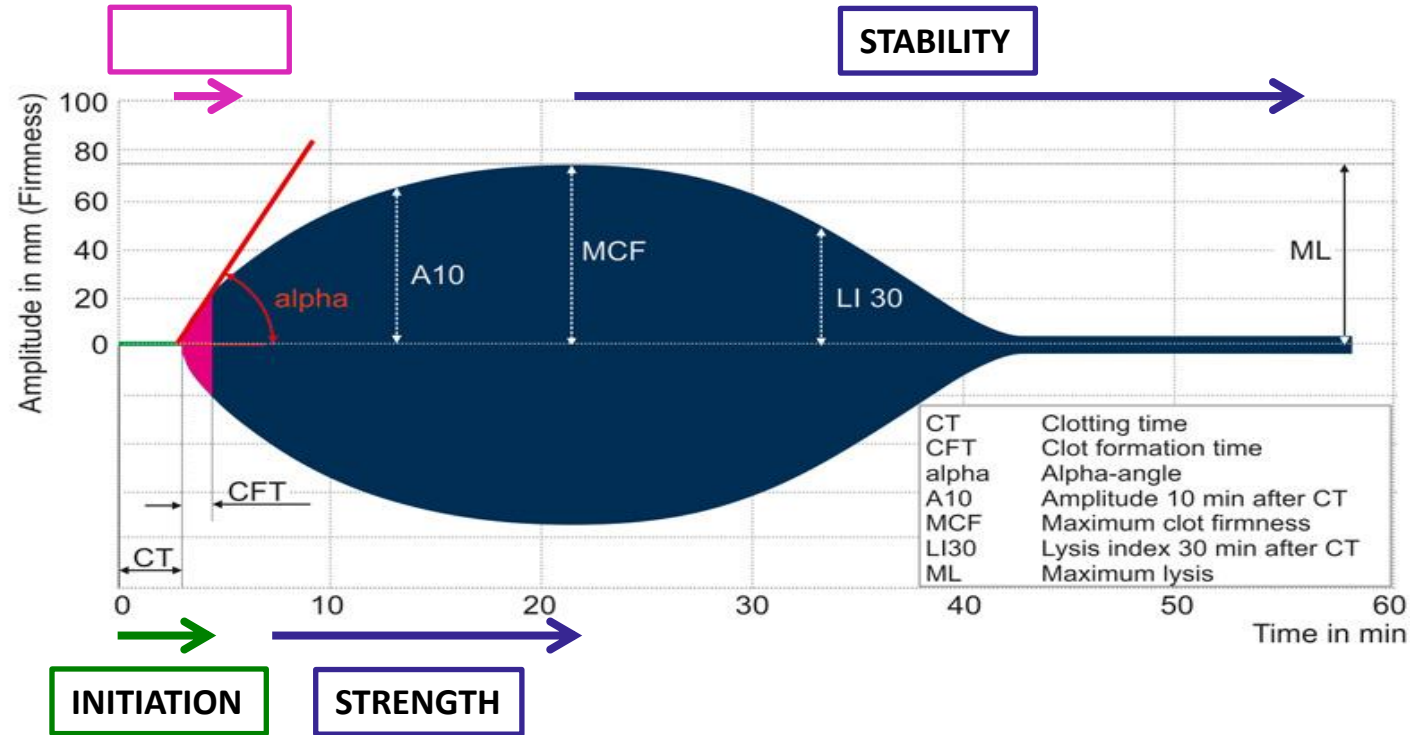
Figure 2. EXTEM and FIBTEM Analysis

Figure 3. GCUH ROTEM Guided Fibrinogen Replacement

Figure 4. Gunshot Wound Abdomen

Figure 5. FEISTY Randomisation and Intervention Flow Chart

Figure



PHASE	ROTEM Parameter	Coagulation Factor
INITIATION	CT - Clotting Time Measured in Seconds	Pro-Coagulant Factors Anti-Coagulant Factors
KINETICS	CFT - Clot Formation Time Measured in Seconds	Pro-Coagulant Factors Anti-Coagulant Factors Fibrinogen Platelets
STRENGTH	Amplitude after CT Measured in mm A5 - Amplitude after 5 mins A10 - Amplitude after 10 mins MCF - Maximum Clot Firmness	Fibrinogen Platelets FXIII
STABILITY	LI (Lysis Index/Residual Clot Firmness) Measured in % of MCF LI30 - 30 mins after CT ML - Maximum Lysis in % of MCF	Fibrinolytic Factors Fibrinolytic Inhibitors

Figure 2



EXTEM			
CT:	67s	CFT:	87s
CFR:	54mm	MCF:	57mm
		ML:	-%

EXTEM:

Activator: CaCl_2 + recombinant Tissue Factor

CT (Clotting Time) - Clotting Factors

Clot Amplitude (A5/A10/MCF) - Interaction of Fibrinogen and Platelets

↓ Clot Amplitude **EXTEM** + Normal **FIBTEM** = Poor Platelet Contribution

↑ CT + Normal **FIBTEM** = Factor Deficiency or Anticoagulants



FIBTEM			
CT:	66s	CFT:	-s
CFR:	9mm	MCF:	10mm
		ML:	-%

FIBTEM:

Activator: CaCl_2 + recombinant Tissue Factor

Inhibition of Platelet Contribution by addition of Cytochalasin D

Clot Amplitude (A5/A10/MCF) - Fibrinogen Contribution

↓ Clot Amplitude **FIBTEM** = Fibrinogen Deficit or ↓ Fib Polymerisation

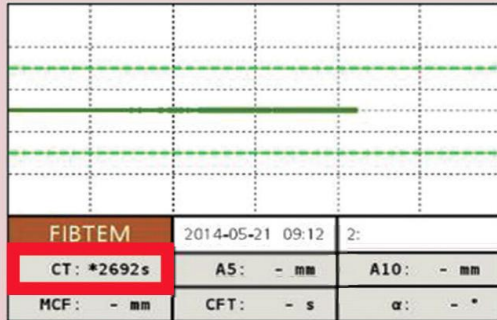
CRITICAL BLEEDING ROTEM TRANSFUSION ALGORITHM

GOLD COAST UNIVERSITY HOSPITAL

Physiological Targets: Temp >36°C pH >7.2 iCa >1 mmol/L Hb>70g/L

STEP 1: HYPERFIBRINOLYSIS

1



FIBTEM CT > 600 sec

AND

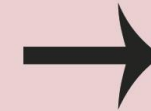
EXTEM A5 < 35 mm



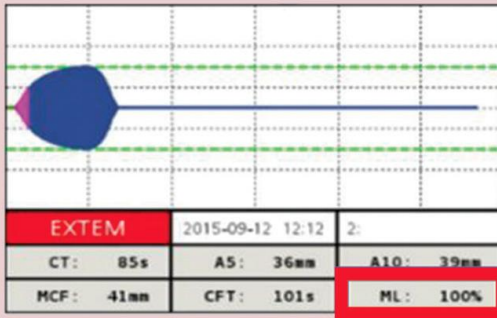
TXA 1g +
FIB CONC 4g

OR

ML% > 5%



TXA
1g

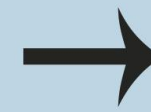


STEP 2: FIBRINOGEN

2



FIBTEM A5 ≤ 8 mm



FIB CONC
1g/25Kg BW

OR

FIBTEM A5 ≤ 10 mm

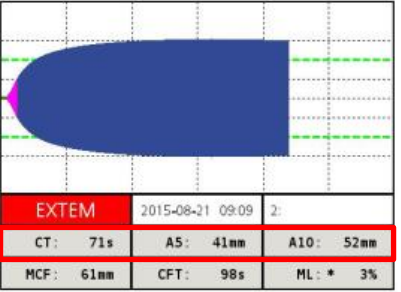
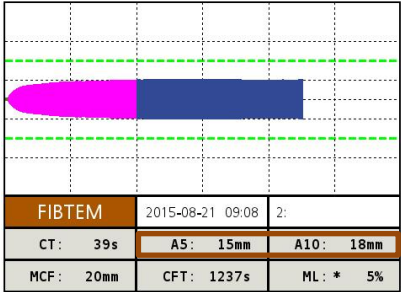


CRYO
1 Unit/5Kg BW

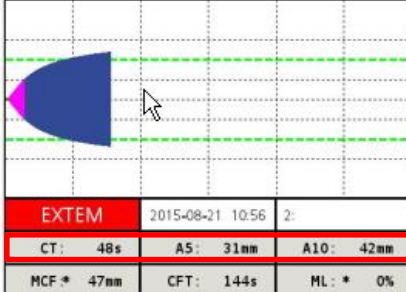
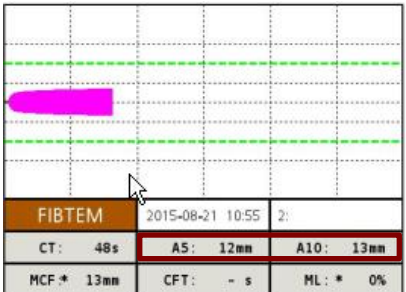
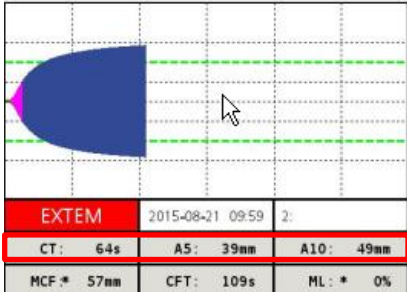
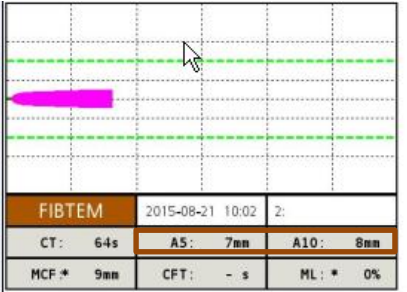
Figure 4

Fig 4. GSW Abdomen

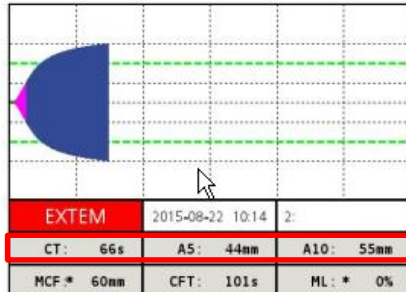
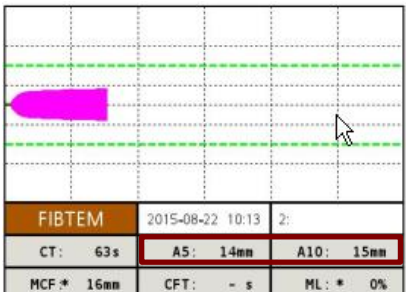
ED Arrival



Operating Theatre



D1 ICU



ED Arrival: Hypotensive, FAST +ve, Lactate 9, BE -14 but Normal ROTEM and Standard Coagulation Tests
In OT: Visceral, vascular and renal tract injuries with significant haemorrhage
DCS: Abdomen Packed + Left Open
Blood product administration guided by ROTEM and utilising FC
ABG at end of case: pH 7.3, Lactate 3, BE -6
Total Transfusion 1st 24hrs: 8 PRBC, 4g FC, 10 Units Cryo, 1 Platelet, 1g TXA
SLT D1: Hb 104, PT 16, Plt 130, Fib 3
Definitive surgery completed 72 hours after admission